

Interaction of amyloid- β protein with raft-model liposome under mimic environment of cellMitsuhiro Hirai^{1,*}, Shoki Sato¹, Nobutaka Shimizu², and Noriyuki Igarashi²¹ Graduate School of Engineering, Gunma University, Maebashi, Gunma 371-8510, Japan.² Photon Factory, Tsukuba 305-0801, Japan

1 Introduction

It is now widely accepted that the amyloid β -peptide ($A\beta$), which exists in fibrillar forms as a major component of senile plaques, is central to the development of Alzheimer's disease (AD). $A\beta$ -membrane interactions have been extensively investigated to elucidate the molecular mechanisms of the $A\beta$ -induced cellular dysfunctions underlying the pathogenesis of AD. Particularly, according to accumulating evidence using spectroscopic methods, the importance of ganglioside clusters in the fibrillization of $A\beta$ is emphasized. Gangliosides, major components of glycosphingolipids (GSLs), are acidic lipids and rich in central nervous system. GSL-cholesterol formed microdomains in cell membranes, so-called lipid rafts, are suggested to function as platforms of various membrane-associated events such as signal transduction, cell adhesion, lipid/protein sorting and so on. The interaction between mono-sialoganglioside (G_{M1}) and $A\beta$ is reported to promote the conformational change of $A\beta$ to its cross-beta structure and the G_{M1} -bound form seeds in the process of $A\beta$ polymerization to amyloid fibril [1]. Recently, we reported that the interaction between the $A\beta$ proteins and the model membrane at the liquid-crystal phase significantly suppresses a bending-diffusion motion, suggesting the possibility of non-receptor-mediated disorder in signaling through a modulation of a membrane dynamics induced by the association of amyloidogenic peptides on a plasma membrane [2].

2. Experimental

In the present study, we have investigated the $A\beta$ -membrane interactions under a mimic intracellular environment by using the time-resolved small- and wide-angle X-ray scattering (SWAXS) by using the synchrotron radiation small-angle X-ray scattering spectrometer installed at BL10C beam port of PF at KEK, Tsukuba, Japan. The X-ray wavelengths were 1.0 Å and 1.5 Å. The sample-to-detector distances were 50 cm and 300 cm. X-ray scattering intensity was recorded by a hybrid pixel detector, PILATUS3 2M (Dectris). The exposure time was 60 seconds. Sample cells composed of a pair of thin-quartz windows with 1 mm path length were used. The membrane used was the small unilamellar vesicle (SUV) composed of G_{M1} , cholesterol, and phospholipid. The molar ratio [G_{M1}]/[cholesterol]/[DPPC or DOPC] of the mixtures was 0.1/0.1/1, where the phospholipid concentration was fixed at 1 % w/v. Amyloid β protein (1-40) purchased from Wako Chem. Co. was used without further purification. The solvent

condition was 10mM Hepes, pH7.4, 50mM NaCl, 5mM KCl, 2mM $MgCl_2$, 2mM $CaCl_2$. The mimic intracellular environment, so-called crowding environment, was realized by the addition of the high-molecular weight neutral polymer (polyvinyl-pyrrolidone, PVP).

3. Results and Discussion

Fig. 1 shows the time evolution of SWAXS curve of the SUV (G_{M1} -cholesterol-DPPC) after adding $A\beta$ protein. The present results are summarized as follows. Under non-crowding environment, the interaction between SUV and $A\beta$ protein proceeds mostly without affecting the SUV membrane structure. While, under crowding environment, the presence of crowders induces the deformation of the SUV shape and the interaction between SUV, and $A\beta$ protein causes further aggregation to form the lamellar stacking. The time-course of the lamellar distance and the lamellar-peak height suggest the preferential cohesion or intercalation of $A\beta$ proteins in the inter-bilayer region.

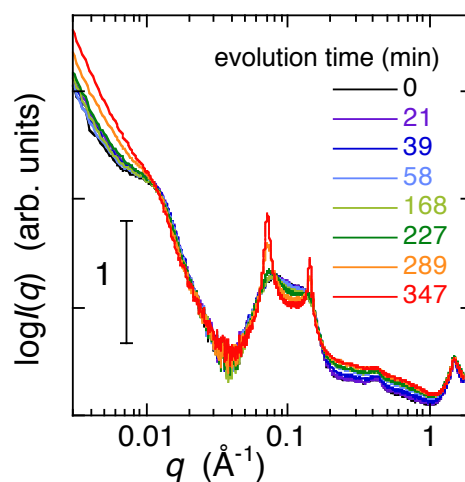


Figure 1. Time-evolution of SAWXS curve of SUV after $A\beta$ addition

References

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* mhirai@gunma-u.ac.jp