

# Crystallographic analysis of diadenosine tetraphosphate phosphorylase from *Mycobacterium tuberculosis* H37Rv complexed with ADP

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## 1 Introduction

Recently, we found that Rv2613c protein of *Mycobacterium tuberculosis* H37Rv was a novel diadenosine tetraphosphate (Ap<sub>4</sub>A) phosphorylase and determined its crystal structure [1,2]. Results of structure–function analyses suggested that *M. tuberculosis* Ap<sub>4</sub>A phosphorylase had a unique substrate-binding site. In the present study, we obtained crystals of *M. tuberculosis* Ap<sub>4</sub>A phosphorylase complexed with ADP to assess its substrate-binding site in detail.

## 2 Experiment

*M. tuberculosis* Ap<sub>4</sub>A phosphorylase was expressed and purified, as described previously [1]. Next, *M. tuberculosis* Ap<sub>4</sub>A phosphorylase complexed with ADP was crystallized by performing a hanging drop vapor diffusion method in a 24-well plate. Briefly, 5  $\mu$ L protein solution (10 mg·mL<sup>-1</sup>) containing ADP (0.1 mM) was mixed with 5  $\mu$ L crystallization solution (0.1 M sodium cacodylate [pH 6.3], 0.2 M lithium sulfate, and 28.5% polyethylene glycol 400). A drop of this mixture was suspended on a siliconized coverslip over 0.6 mL crystallization solution. Prismatic colorless crystals of *M. tuberculosis* Ap<sub>4</sub>A phosphorylase complexed with ADP developed after approximately 2 weeks of incubating the drop at 20°C and grew to a maximum size of 0.3 mm. Diffraction data of *M. tuberculosis* Ap<sub>4</sub>A phosphorylase complexed with ADP were collected at BL-5A, BL-17A, and AR-NW12A stations. The crystals were harvested using a cryoloop, were flash-cooled to 100 K under nitrogen stream, and were analyzed. Next, 360 consecutive images were collected using an oscillation range of 1°, exposure time of 2 s, and wavelength of 1.0000 Å. The diffraction data were processed, merged, and scaled using HKL2000 (DENZO and SCALEPACK) software package. Values of Wilson B-factor, Matthew's coefficient, and solvent content were calculated using CCP4 program.

## 3 Results and Discussion

The diffraction data of crystals of *M. tuberculosis* Ap<sub>4</sub>A phosphorylase complexed with ADP were collected using synchrotron radiation. A single crystal resulted in a diffraction of 3.2 Å. Preliminary characterization of the crystal indicated that it belonged to monoclinic space group C2, with unit cell parameters of  $a = 101.7$ ,  $b = 63.4$ ,  $c = 79.3$  Å, and  $\beta = 111.2^\circ$ . Furthermore, 7,462 independent reflections were obtained using an  $R_{\text{merge}}$  of 15.9%. Data collection statistics for the crystal are summarized in Table 1.

*M. tuberculosis* Ap<sub>4</sub>A phosphorylase formed a homotetramer of 25-kDa subunits in solution [1,2]. One asymmetric unit of *M. tuberculosis* Ap<sub>4</sub>A phosphorylase included two subunits, with a Matthew's coefficient of 2.38 Å<sup>3</sup>·Da<sup>-1</sup> and solvent content of 48.43%.

Studies assessing the structure of *M. tuberculosis* Ap<sub>4</sub>A phosphorylase complexed with ADP by using molecular replacement method are in progress. Coordinates of the structure of wild-type *M. tuberculosis* Ap<sub>4</sub>A phosphorylase (PDB code: 3ANO) were used as a search model in the present study.

Table 1: Summary of data collection and processing

Wavelength (Å)	1.00000
Temperature (K)	100
Space group	C2
Cell dimension ( $a, b, c$ ) (Å)	101.7, 63.4, 79.3
$\beta$ (deg)	111.2
Resolution range (Å)	73.94–3.20 (3.37–3.20) <sup>a</sup>
Total No. of reflections	49961 (7247)
No. of Unique reflections	7462 (1068)
Redundancy	6.7 (6.8)
Completeness (%)	95.1 (94.9)
Mean $I/\sigma(I)$	10.0 (4.5)
$R_{\text{merge}}^b$	15.9 (40.2)
Wilson B-factor (Å)	24.4

<sup>a</sup>Data in the highest resolution shells are listed in parentheses.

<sup>b</sup> $R_{\text{merge}} = \frac{\sum_{hkl} \sum_i |I_i(hkl) - \langle I(hkl) \rangle|}{\sum_{hkl} \sum_i I_i(hkl)}$ , where  $I_i(hkl)$  is the  $i$ th intensity measurement of reflection  $hkl$  and  $\langle I(hkl) \rangle$  is its average.

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## References

- [1] S. Mori *et al.*, Protein. Expr. Purif. **69** (2010) 99-105.  
[2] S. Mori *et al.*, J. Mol. Biol. **410** (2011) 93-104.

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