

Crystal structure of 1,4- $\beta$ -mannanase from *Eisenia fetida*Yu Hirano<sup>1</sup>, Mitsuhiro Ueda<sup>2</sup>, Taro Tamada<sup>1\*</sup><sup>1</sup>National Institutes for Quantum and Radiological Science and Technology, Shirakata 2-4, Tokai, 319-1116, Japan<sup>2</sup>Osaka Prefecture University, 1-1 Gakuen-cho, Sakai, Osaka, 599-8531, Japan

### 1 Introduction

Endo-1,4- $\beta$ -mannanase have been used for various bioprocesses such as bleaching of softwood pulps, declining viscosity of feeds and foods, and clarifying beverages. 1,4- $\beta$ -mannanase has been isolated from bacteria, fungi, higher plants, mollusks, and arthropods, but it has not been reported about mannanase from annelida earthworm.

We have found a high mannanase activity in the body wall extract of the earthworm *Eisenia fetida*. We have cloned and expressed 1,4- $\beta$ -mannanase from *Eisenia fetida* (Ef-Man). To understand the structure and function of Ef-Man, we have determined crystal structures of Ef-Man.

### 2 Experiment

Ef-Man crystals were obtained under the conditions containing ammonium citrate (crystal-1) or sodium potassium phosphate (crystal-2) as precipitants. Crystals (crystal-2) were soaked in 10 mM mannitriose (Man3) for 1 day. The crystal-1 and crystal-2 were transferred to the cryo-protectant solution consisting of the mother liquor with 20% glycerol. The crystals were cooled in a nitrogen-gas stream at 100 K during X-ray data collection.

Diffraction data sets were collected on the BL-17A beamline. The wavelength of X-rays was set to 0.98 Å and diffraction intensities were measured with Pilatus3 S6M detector. Crystal structures of Ef-Man were determined by the molecular replacement method with the crystal structure of 1,4- $\beta$ -mannanase from *Mytilus edulis* (PDB 2C0H) [1] as a search model.

### 3 Results and Discussion

Thin plate crystals were obtained in both crystallization conditions. The crystal-1 and crystal-2 were diffracted to 1.7 Å and 1.6 Å resolutions, respectively (Table 1). Both crystals belong to the space group  $P2_12_12_1$  with unit cell parameters of  $a = 50.1$  Å,  $b = 69.5$  Å,  $c = 86.6$  Å (crystal-1) and  $a = 50.0$  Å,  $b = 69.0$  Å,  $c = 85.5$  Å (crystal-2).

The final models of Ef-Man consist of Gln17-Gln377. The overall structures of crystal-1 and crystal-2 show high similarities each other. The superposition of the two structures indicated the root-mean-square distance of 0.18 Å for all C $\alpha$  atoms. The crystal structure of Ef-Man also shows high similarities to the crystal structures of 1,4- $\beta$ -mannanase from other species. Superposition of the Ef-Man structure to the mannanase structures shows rms distances of 1.04 Å for 347 C $\alpha$  atoms (*M. edulis*, 2C0H), 0.87 Å for 357 C $\alpha$  atoms (*Cryptococcus antarcticus*,

400U) [2], and 0.90 Å for 329 C $\alpha$  atoms (*Aplysia kurodai*, 3VUP) [3].

In the crystal-1 structure of Ef-Man, one Tris molecule that included in the crystallization solution was bound to the active site. Although crystal-2 was soaked in the Man3 solution, there are no clear density of Man3 at the active site of the crystal-2 structure. Instead of Man3, water molecules were bound to the active site (Fig. 1). To obtain the structure complexed with Man3, we will modify soaking condition or perform co-crystallization with substrates.

Table 1: Diffraction data statistics

	crystal-1	crystal-2
Beamline	BL-17A	BL-17A
Wavelength (Å)	0.98	0.98
Resolution (Å)	50-1.70 (1.81-1.70)	50-1.60 (1.70-1.60)
Unique reflections	62 976	74 549
Completeness (%)	98.4 (97.1)	99.4 (98.6)
$R_{\text{sym}}$ (%)	7.4 (46.4)	9.7 (52.7)
$\langle I \rangle / \langle \sigma(I) \rangle$	13.1 (2.7)	10.3 (3.0)

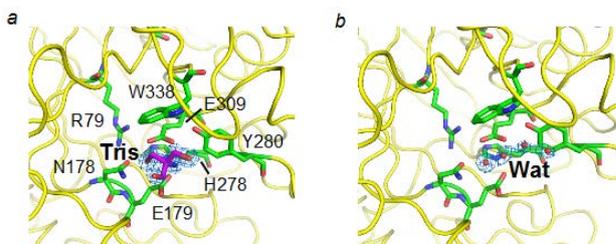


Fig. 1: Electron density at the active site of Ef-Man.  $2F_o - F_c$  map (blue mesh) of (a) crystal-1, (b) crystal-2.

### Acknowledgement

We thank the PF beamline staff for data collection.

### References

- [1] A.M. Larsson *et al.*, *JMB* **357**, 1500 (2006).
- [2] M.K. Kim *et al.*, *Proteins* **82**, 3217 (2014).
- [3] K. Mizutani *et al.*, *Acta Cryst. F* **68**, 1164 (2012).

\* tamada.taro@qst.go.jp