## Sequence-Selective Recognition of Peptides within the Single Binding Pocket of a Self-Assembled Coordination Cage

Shohei Tashiro,<sup>1</sup> Masahide Tominaga,<sup>1</sup> Masaki Kawano,<sup>1</sup> Bruno Therrien,<sup>1</sup> Tomoji Ozeki,<sup>2</sup> and Makoto Fujita<sup>1</sup>\*

<sup>1</sup> Department of Applied Chemistry, The University of Tokyo, CREST, Japan Science and Technology Corporation (JST), Bunkyo-ku, Tokyo 113-8656, Japan

<sup>2</sup> Department of Chemistry and Materials Science, Tokyo Institute of Technology, Meguro-ku,

Tokyo 152-8551, Japan

## **Introduction**

Sequence-selective recognition of peptides is expected to be an essential process for the site-specific recognition of protein surfaces, which leads to the control of protein functions and to the understanding of biological events at protein surfaces such as protein-protein or proteinoligopeptide interactions. Although a few groups have reported artificial receptors for sequence-selective peptide recognition, the highly designed receptors are in their early stages. Here, we report that the single binding pocket of self-assembled coordination cage 1 can accommodate oligopeptides in a highly sequenceselective fashion. Having a large hydrophobic cavity, cage 1 binds as many as three amino acid residues. X-ray analyses reveal that the sequence-selective recognition is ascribed to cooperative multiple interactions between the residues and the cavity.<sup>1</sup>



## **Results**

We found that cage 1 bound Ac-Trp-Trp-Ala-NH<sub>2</sub> (2) very strongly ( $K_a > 10^6$  M<sup>-1</sup>). Strong binding was specific to the Trp-Trp-Ala sequence because the binding of tripeptides possessing those same residues in different sequences, such as Ac-Trp-Ala-Trp-NH<sub>2</sub> (3) and Ac-Ala-Trp-Trp-NH<sub>2</sub> (4), was much less effective ( $K_a = 2.5 \times 10^5$  and 2.1 x 10<sup>4</sup> M<sup>-1</sup>, respectively). Even singly mutated tripeptides, such as Ac-Trp-Trp-Gly-NH<sub>2</sub> (5) and Ac-Trp-Tyr-Ala-NH<sub>2</sub> (6), showed poorer affinity ( $K_a = 7.4 \times 10^4$  and 5.3 x 10<sup>4</sup> M<sup>-1</sup>, respectively) although they have very similar aromatic-aromatic-aliphatic sequences. These results suggest that the two indole rings and the Ala



Figure 1. Crystal structure of 1.2. Peptide 2 in the cavity is represented by (a) space-filling and (b) cylindrical model. The  $\pi$ - $\pi$  interactions of 1 with indole rings of (c) W1 and (d) W2. (e) The CH- $\pi$  interaction between 1 and methyl group of A3.

methyl group in 2 should cooperatively interact with the cage in the 1.2 complex.

In fact, the multiple interactions of the methyl and indole groups with the cage were revealed by X-ray crystallographic analysis. Single crystals were obtained after an aqueous solution of 1.2 complex stood at room temperature for 4 d. The diffraction data were collected by synchrotron X-ray irradiation. The crystallographic analysis showed that tripeptide 2 is fully encapsulated in the cavity of 1 (Figure 1a,b). As predicted, all residues interact very efficiently with cage 1. Namely, two indole rings are stacked on the triazine ligand by  $\pi$ - $\pi$  interaction (3.4-3.5 Å), while the Ala methyl group interacts with another ligand by CH- $\pi$  contact (2.5 Å) (Figure 1c-e). Despite the enclathration within the restricted cavity, the peptide backbone is fixed in an extended conformation.

## **References**

[1] S. Tashiro et. al. J. Am. Chem. Soc., 127, 4546 (2005).

\*mfujita@appchem.t.u-tokyo.ac.jp