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Crystal Structure Analysis of Enzymes Involved in Biosynthesis of Marihuana Compounds

Taro TAMADA¹, Yoshinari SHOYAMA¹, Chiho TAGUCHI¹, Futoshi TAURA², Ryota KUROKI¹, Satoshi MORIMOTO^{*2} ¹JAEA-QuBS, Shirakata-Shirane, Tokai, Ibaraki 319-1195, Japan

²Kyushu Univ., Maidashi, Higashi-ku, Fukuoka 812-8582, Japan

Introduction

Cannabinoids, which are produced predominantly in *Cannabis sativa*, are unique secondary metabolites consisting of alkylresorcinol and monoterpene groups. More than 60 cannabinoids have been isolated from marijuana or fresh *Cannabis* leaves, and their biosynthetic pathways have been extensively investigated.

We demonstrated that the major cannabinoids (tetrahydrocannabinolic acid and cannabidiolic acid) are biosynthesized by novel FAD-dependent oxidases named tetrahydrocannabinolic acid synthase (THCA synthase) [1] and cannabidiolic acid synthase (CBDA synthase) [2], respectively, from the common precursor cannabigerolic acid which is produced from olivetolic-acid (OLA) and geranylpyrophosphate. We have recently cloned the gene coding for a plant type III polyketide synthase, named polyketide synthase-1 (PKS-1), which catalyzes OLA production. Here we report the X-ray diffraction studies of THCA synthase and PKS-1.

THCA synthase

Single crystals of THCA synthase suitable for X-ray diffraction measurement with the dimension of 0.1×0.1× 0.1 mm were obtained in 0.09 M HEPES buffer (pH 7.5) containing 1.26 M sodium citrate [3]. This crystal belonged to the primitive cubic space group P432, with unit-cell parameters a=b=c=178.2 Å. The calculated Matthews coefficient was approximately 4.1 Å³ Da⁻¹ assuming the presence of one molecule of THCA synthase in the asymmetric unit. Diffraction data was collected at NW12A in PF-AR, and was completed up to 2.6 Å resolution with completeness of 97.5% and R_{merge} of 0.99. The structure was solved by molecular replacement using the coordinates of glucooligosaccharide oxidase (PDB ID: 2AXR). The final model of the THCA synthase (residues 28-545) containing a FAD molecule, five N-linked sugar chains, and 43 waters was refined to a $R_{\text{cryst}}/R_{\text{free}}$ of 0.207/0.259 at 2.6 Å resolution.

Overall structure of THCA synthase is shown in Figure 1. The polypeptide chain of THCA synthase is folded into 14 α -helices, six short 3₁₀-helices, and 16 β -strands. THCA synthase consists of two domains. The FAD binding domain (residues 28-253 and 476-545) folds into two $\alpha + \beta$ subdomains. The FAD molecule is positioned between two subdomains (colored by blue and magenta in Figure 1). The isoalloxazine ring of FAD is cross-linked

to the enzyme at two sites (His114 and Cys176). The other domain is colored by green in Figure 1. This domain (residues 254-475) is composed of an antiparallel β sheet and several helices. Crystallization of substrate complex is now in progress.

<u>PKS-1</u>

The diffraction quality crystals of PKS-1 were obtained under the condition, 100 mM HEPES buffer (pH 7.5) containing 200 mM calcium acetate and 20% (w/v) PEG 3350 [4]. The typical crystal size of each condition was approximately 0.02×0.02×0.3 mm. This Crystal belonged to space group P1 with the unit-cell parameters of a = 54.3, b = 59.3, c = 62.6 Å, $\alpha = 69^{\circ}$, $\beta = 81^{\circ}$, $\gamma = 80^{\circ}$. The calculated Matthews coefficient was approximately $2.2 \text{ Å}^3 \text{Da}^{-1}$ assuming the presence of two molecules of PKS-1 in the asymmetric unit. Diffraction data was collected at BL-5A in PF, and was completed up to 1.65 Å resolution with completeness of 90.4% and $R_{\rm merge}$ of 0.52. The structure was solved by molecular replacement using the coordinates of chalcone synthase (PDB ID: 1BI5). Crystallographic refinement is now in progress.



Figure 1. Overall structure of THCA synthase. FAD molecule is indicated as stick model.

References

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* morimoto@phar.kyushu-u.ac.jp