

## Crystal structural analysis of the complex of tRNA<sup>Asp</sup> mutant and Arginyl-tRNA synthetase from *Pyrococcus horikoshii*

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### Introduction

The aminoacyl-tRNA synthetases exhibit *in vitro* fidelity of one error per 10<sup>4</sup> to 10<sup>5</sup> codons. Surprisingly, it was reported [1] that *Saccharomyces cerevisiae* arginyl-tRNA synthetase (ArgRS) *in vitro* arginylates a non-cognate tRNA<sup>Asp</sup>(GUC) transcript with about 10 fold less efficiency than a cognate tRNA<sup>Arg</sup>(UCU) transcript and further that the  $k_{cat}/K_m$  value of tRNA<sup>Asp</sup> U34 mutant is the same as that of tRNA<sup>Arg</sup>. These facts reveal that in the anticodon-binding domain of ArgRS, the base U34 of tRNA<sup>Asp</sup>(UUC) mutant binds at the site accepting U34 of tRNA<sup>Arg</sup>(UCU). The fact that tRNA<sup>Met</sup>(CAU) is also arginylated by *E. coli* ArgRS [2] expects that the bases of the anticodon C34, A35, U36 interact with sites accepting C34, C35 and G36 bases of tRNA<sup>Arg</sup>, respectively. Taking these reports into consideration, the bases of U35 and C36 of tRNA<sup>Asp</sup>(UUC) mutant also interact with sites accepting 35<sup>th</sup> and 36<sup>th</sup> bases, respectively. tRNA<sup>Asp</sup>(GUC) is not common to the second base C35 and the third base G36/U36 of the anticodon of tRNA<sup>Arg</sup> but this paper [1] proposed 'slipped' recognition set hypothesis that in *S. cerevisiae* tRNA<sup>Asp</sup>(GUC), C36-G37 may be used in place of C35-G36 of tRNA<sup>Arg</sup>. The relative orientation of G37 base for C36 base in *S. cerevisiae* tRNA<sup>Asp</sup>(GUC) [3] is quite different from the relative orientation of G36/U36 base for C35 base in *S. cerevisiae* tRNA<sup>Arg</sup>(ICG) [4] and *Pyrococcus horikoshii* tRNA<sup>Arg</sup>(CCU) [5] (Figure 1). Therefore, the 'slipped' recognition set hypothesis is less possible. In order to prove that bases of the anticodons of tRNA<sup>Asp</sup> mutants interact with sites accepting C34, C35, U36 bases of tRNA<sup>Arg</sup>, we tried crystal structure analysis of the complex of *P. horikoshii* ArgRS and tRNA<sup>Asp</sup> mutants.

### Results

#### Experiment

The D loop of all *P. horikoshii* tRNA<sup>Asp</sup> mutants was substituted by that (AGCCA<sup>17a</sup>GGAC<sup>20a</sup>A) of *P. horikoshii* tRNA<sup>Arg</sup>(CCU) such that tRNA<sup>Asp</sup> does not occur steric obstacle with ArgRS. The nucleotides of anticodon loop of tRNA<sup>Asp</sup> are CUGUCAC, whereas those of tRNA<sup>Arg</sup>(CCU) are CUCCUAA. The genes of tRNA<sup>Asp</sup> mutants with the anticodon of UUC, GUC, CCU were cloned with the T7 promoter into the vector pUC119. The crude transcript was purified by Resource Q column. The crystallizations of complexes of ArgRS and tRNA<sup>Asp</sup> mutants were tried at the same condition as that of the

complex of tRNA<sup>Arg</sup>. Crystals of the complex of tRNA<sup>Asp</sup>(CCU) hybrid containing the tRNA<sup>Arg</sup> body and the tRNA<sup>Arg</sup> anticodon loop grew. The crystal was exposed at 100 K on beamline AR-NE3A at the Photon Factory. The reflections were observed at 4.5 Å resolution but not enough to analyze.

We also constructed the model of tRNA<sup>Asp</sup>(GUC) and its mutants, C34-C35-U36, U34-U35-C36 on the basis of the structures of complexes of tRNA<sup>Arg</sup>(ICG) and *S. cerevisiae* ArgRS [4] and of tRNA<sup>Arg</sup>(CCU) and *P. horikoshii* ArgRS [5] (Figure 1). The anticodon stem and the D stem of tRNA<sup>Asp</sup>(GUC) [3] were superimposed into those of *S. cerevisiae* tRNA<sup>Arg</sup>(ICG) bound a ArgRS. Then the backbone of the anticodon loop of tRNA<sup>Arg</sup>(ICG) is kept and the bases are replaced by those of tRNA<sup>Asp</sup>(GUC). U35 and C36 are inserted into hydrophobic pockets accepting C35 and G/U36 in ArgRS, respectively.

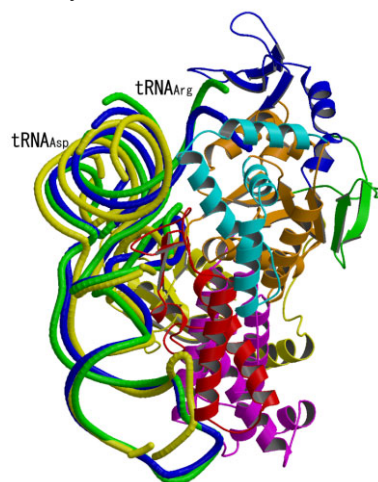


Figure 1. Model of tRNA<sup>Asp</sup>(GUC) (thin) [3] and tRNA<sup>Arg</sup>(ICG) (thick) [4] superimposed into tRNA<sup>Arg</sup>(CCU) (thin) bound *P. horikoshii* ArgRS [5]

### References

- [1] M. Sissler, R. Giegé, & C. Florentz, *EMBO J*, **15**, 5069-5076, 1996.
- [2] L.H.Schulman & H.Pelka, *Science* **241**, 1595-1597, 1989.
- [3] M. Ruff *et al.*, *Science*, **252**, 1682-1689, 1991.
- [4] B. Delagoutte, D. Moras & J. Cavarelli, *EMBO J*, **19**, 5599-5610, 2000.
- [5] M. Konno *et al.*, *FEBS J*, **276**, 4763-4779, 2009.

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