

X-ray structure of *Bacillus pallidus* D-arabinose isomeraseHiromi YOSHIDA<sup>1</sup>, Kosei TAKEDA<sup>1,2</sup>, Ken IZUMORI<sup>2</sup>, Shigehiro KAMITORI\*<sup>1</sup><sup>1</sup>Life Science Research Center and Faculty of Medicine, Kagawa University  
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Miki-cho, Kita-gun, Kagawa 761-0795, Japan**Introduction**

D-Arabinose isomerase (D-AI) catalyzes the isomerization of D-arabinose (aldose) to D-ribulose (ketose). It is able to catalyze the isomerization of L-fucose to L-fuculose and also known as L-fucose isomerase (L-FI). *Bacillus pallidus* (*B. pallidus*) D-AI can catalyze the isomerization of D-psicose to D-altrose, as well as D-arabinose to D-ribulose and L-fucose to L-fuculose. D-psicose and D-altrose are so-called “rare sugars” which exist in small amounts in nature. Since rare sugars have various physiological functions, they have received much attention in the food, agricultural and medicinal industries. *B. pallidus* D-AI is expected to be used for rare sugar production. We have reported the X-ray structures of *B. pallidus* D-AI and its complex with an inhibitor, L-fucitol [1,2].

**Materials and methods**

A crystal of the complex of *B. pallidus* D-AI/L-fucitol was prepared by soaking in 3.0  $\mu$ l of a reservoir solution containing 2.0 M L-fucitol for 48 h. X-ray diffraction data were collected on the NW-12A beam line in the Photon Factory with the wavelength of 1.0  $\text{\AA}$ . The data were processed using the programs HKL2000 and the CCP4 program suite. The initial phases were determined by a molecular replacement method with the program MOLREP in the CCP4 program suite, using the structure of *E. coli* L-FI (PDB code 1FUI) as a model probe. Further modeling was performed with the programs Coot in the CCP4 program suite, and X-fit in the XtalView program system, and the structure was refined using the programs Refmac5 and CNS.

**Results and discussion**

The structure of the subunit of *B. pallidus* D-AI including 23 $\beta$ -strands and 22 $\alpha$ -helices can be divided into three domains. In a crystal, *B. pallidus* D-AI forms a homo-hexamer with a triangular prismatic shape as shown in Figure 1.

The structure of the active site with the bound L-fucitol is shown in Figure 2. Glu342(OE1), Asp366(OD1), Asp366(OD2), and O2 of L-fucitol coordinate Mn<sup>2+</sup> with a planar structure, and O1 of L-fucitol coordinates the axial position. The distance between Glu342(OE2) and C2 of L-fucitol is 3.3  $\text{\AA}$  and H2 is located between them. On the opposite side, Asp366 forms a hydrogen bond with O1 and O2. This structure strongly supports the enediol mechanism, because Glu342 and Asp366 could be

proposed to transfer a proton from C2 to C1 and O1 to O2, acting as acid/base catalysts.

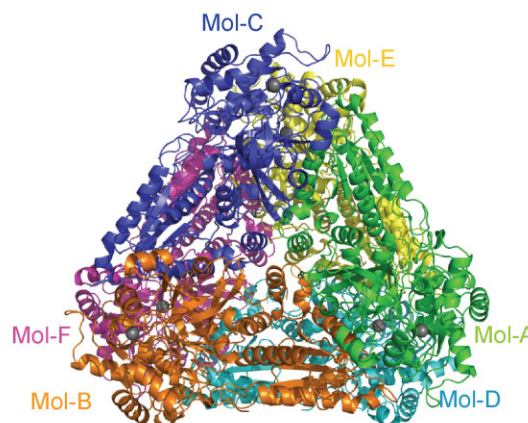


Figure 1. Overall structure of a hexamer of *B. pallidus* D-AI, viewed from two directions. The six molecules are colored in green (Mol-A), orange (Mol-B), blue (Mol-C), cyan (Mol-D), yellow (Mol-E) and magenta (Mol-F).

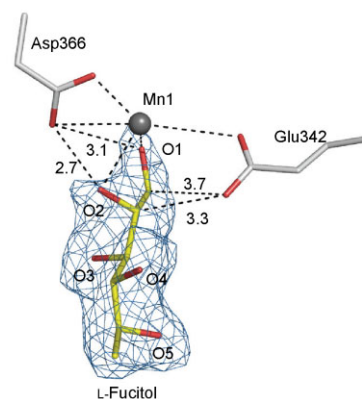


Figure 2. The structure of the active site of *B. pallidus* D-AI in a complex with L-fucitol. Simulated annealing omit maps of the bound ligand are shown at the 4.0  $\sigma$  contour level. Selected interactions among amino acid residues, substrates, metal ions are indicated by dotted lines.

**References**

- [1] K. Takeda *et al.*, Acta Crystallogr. F64, 945-948(2008).
- [2] K. Takeda *et al.*, Biochim. Biophys. Acta. 1804, 1359-1368 (2010).

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