

ERLサイエンスワークショップ
2009.07.10

先端1分子計測と 次世代放射光の役割

Advanced Single Molecular Observations
and the task of Next X-ray Light Sources

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Graduate School of Frontier Sciences
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History of Single Molecular World:

T. Hirschfeld, Applied Optics (1975)

Optical microscopic observation of single small molecules.

A. Ashkin, Phys. Rev. Lett. (1978)

Trapping of Atoms by Resonance Radiation Pressure.

S. Asakura et al., J. Mol. Biol. (1980)

Dark-field Light Microscopic Study of the Flexibility of F-actin Complexes.

G. Binning et al., Phys. Rev. Lett. (1982)

Surface Studies by Scanning Tunneling Microscopy

J. A. Spudich et al., Nature (1983)

Movement of Myosin-coated Fluorescent beads on actin cables in vitro.

T. Yanagida et al., Nature (1984)

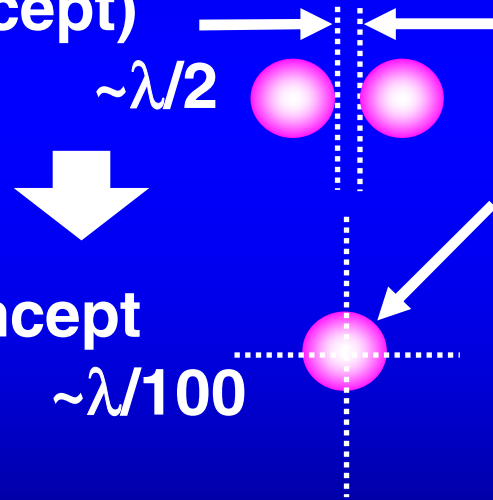
Direct observation of Motion of Single F-actin filaments in the Presence of Myosin.

Y. C. Sasaki et al., Phys. Rev. Lett. (2001)

Picometre-scale Dynamical X-ray Imaging of Single DNA molecules

Positioning Accuracy of Individual Molecules ? from visible light to x-rays

(1) Visible light
($\lambda=300\text{nm}-800\text{nm}$)
Optical Diffraction limit
(imaging concept)



Tracking concept

$\sim\lambda/100$

Nanometer-scale Dynamics

We utilize the tracking concept
in the x-ray regions.

(2) X-ray ($\lambda=1\text{nm}-0.01\text{nm}$)

Tracking $\sim\lambda/100$

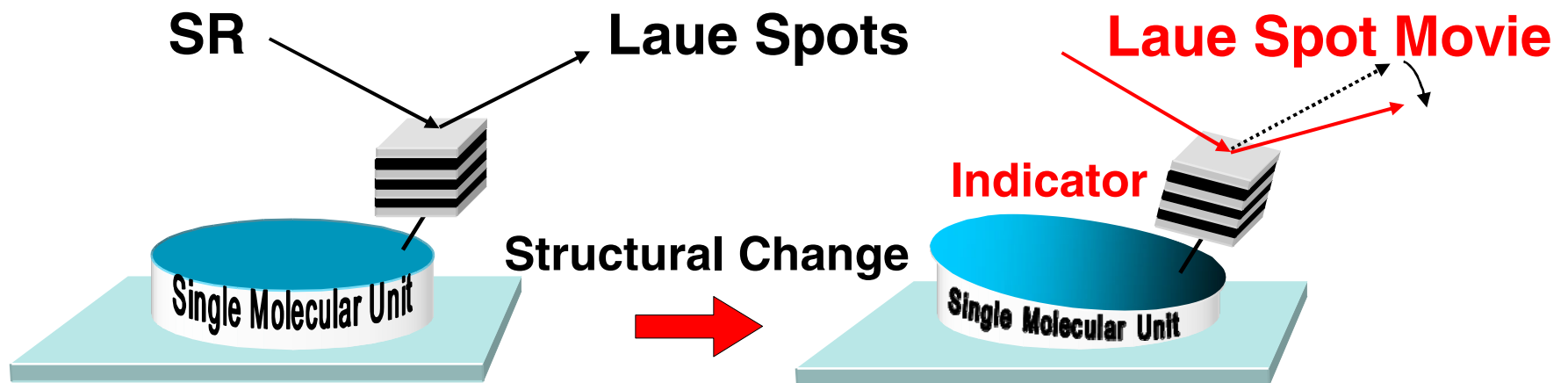
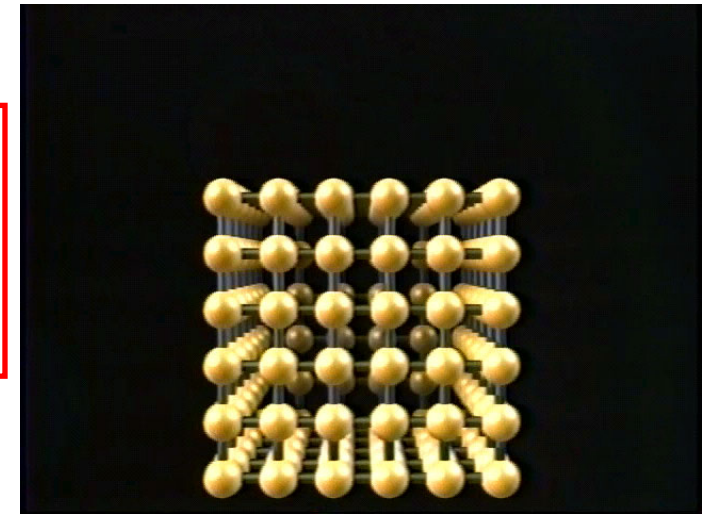
Picometer-scale Dynamics

Principle: Diffracted X-ray Tracking (DXT)

DXT monitors x-ray diffraction from the individual nanocrystal, which is linked to the individual single protein molecule in bio-systems.

Features:

- (1) High Accuracy(-pm=nm/1000)
- (2) Time-resolved Information(from ms to μ s)
- (3) *in vivo* Measurement
- (4) Independent from Chemical Conditions



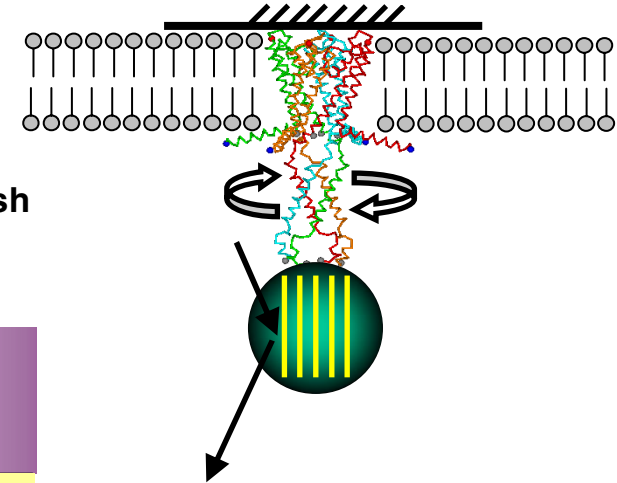
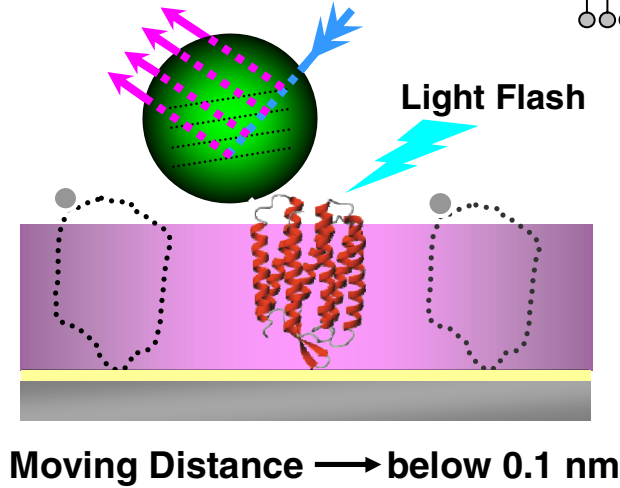
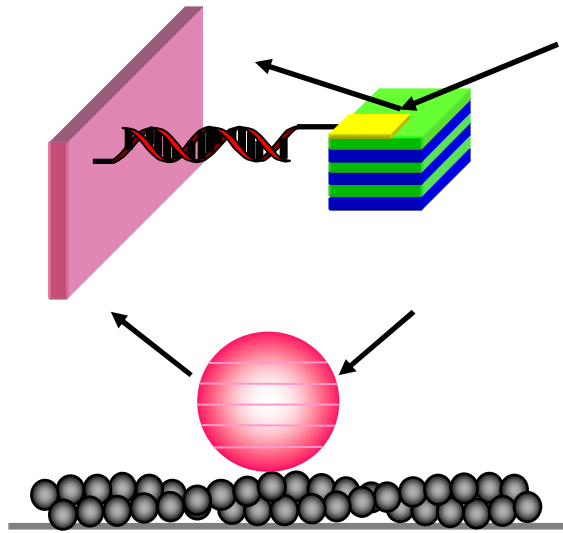
We assumed that motions of a specific site in individual proteins are equal to the observed orientations of nanocrystals.

Single Molecular Measurements (comparison): **Fluorescent, FRET and X-rays (DXT)**

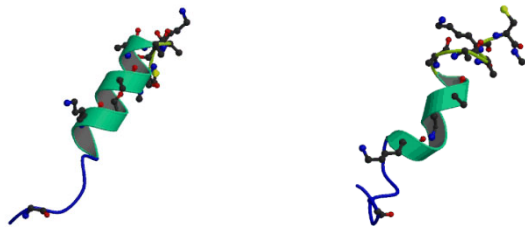
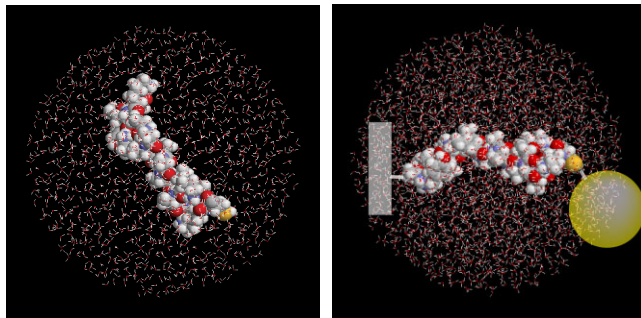
FRET: Fluorescent Resonance Energy Transfer

	Fluorescent	FRET (Electron Transfer)	X-ray (DXT)
Labeled	Molecule Particle	Acceptor/Donor GFP, cy-3, FAD	Nanocrystal
Accuracy	-nm	-nm- 0.1nm	- 0.001nm
Chemical Infor.	Independence	Dependence	Independence
Intramolular Sturactical Infor.	△	○	⊙
(Wavelength)		(300-800 nm)	(0.1-0.01nm)

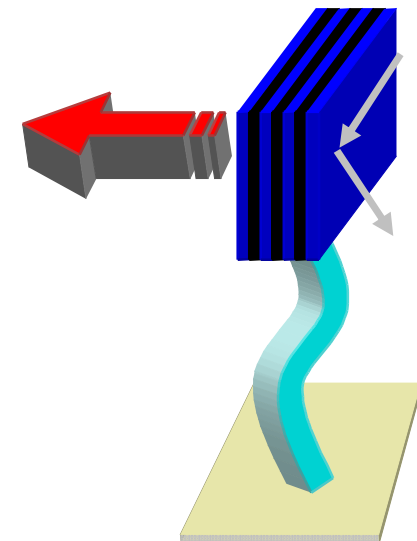
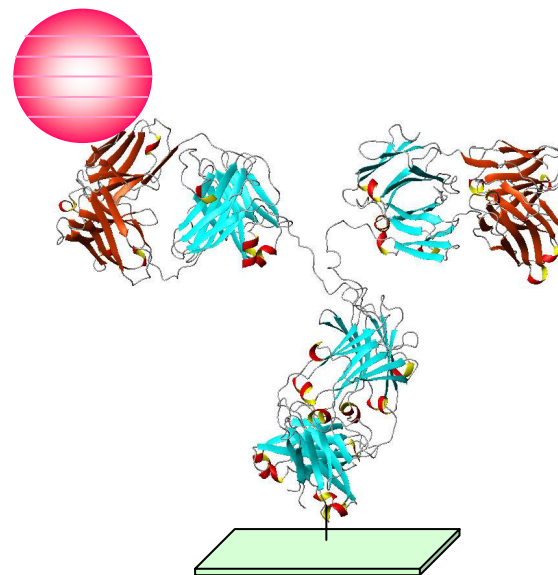
Experimental Samples:



X-ray Radiation Pressure



Replica Exchange Molecular Dynamics Simulation



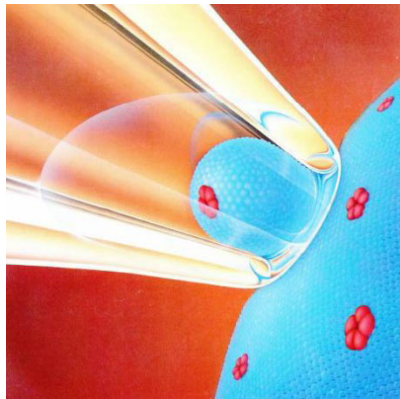
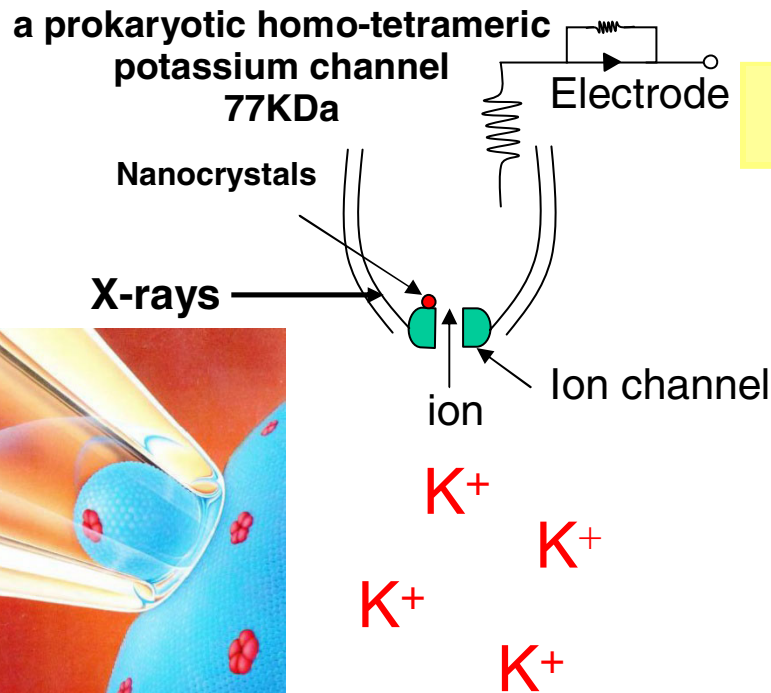
Experiment: Potassium Channel (KcsA)

H. Shimizu, M. Iwamoto, F. Inoue, T. Konno, Y.C. Sasaki, S. Oiki: *Cell* 132, 67 (2008)

Final goal: Simultaneous Observations of Single Molecule

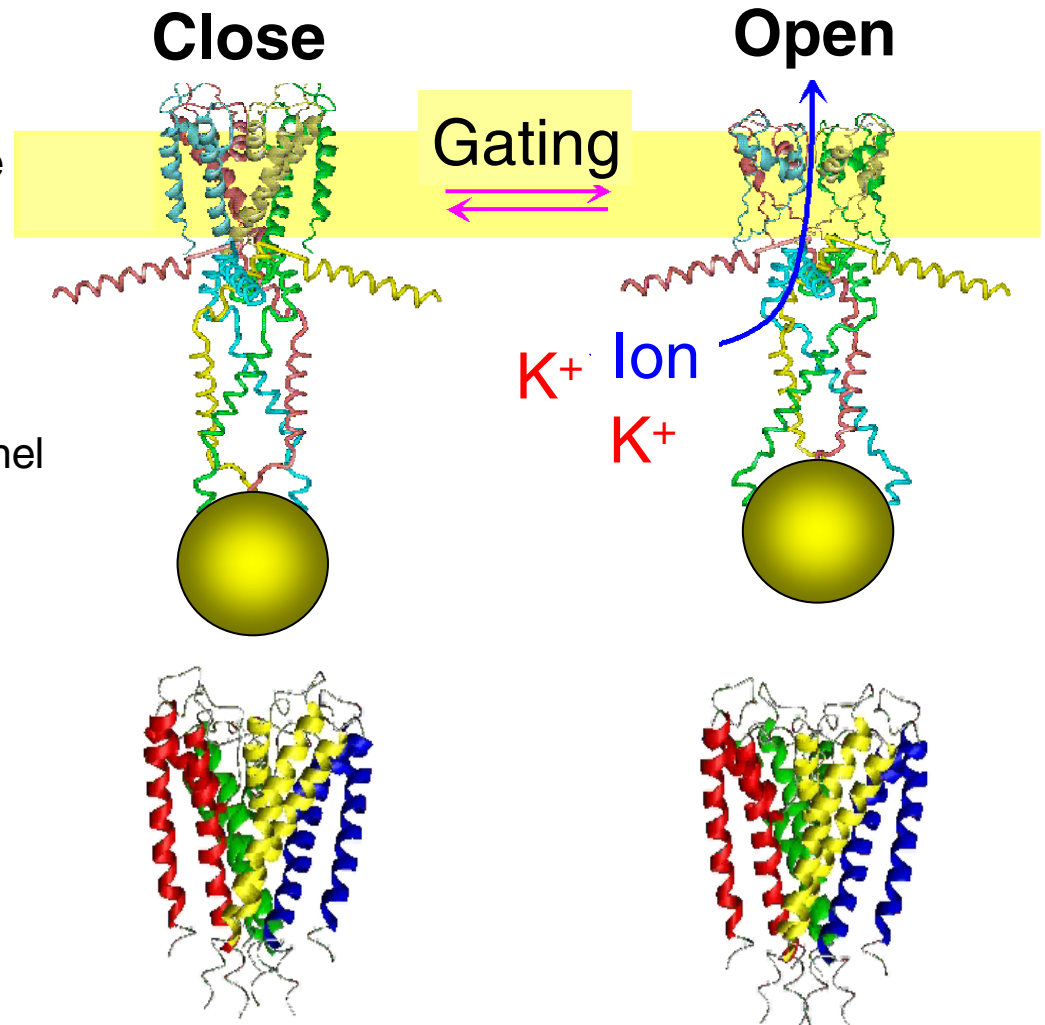
Expected Open and Close-state

Structures of KcsA

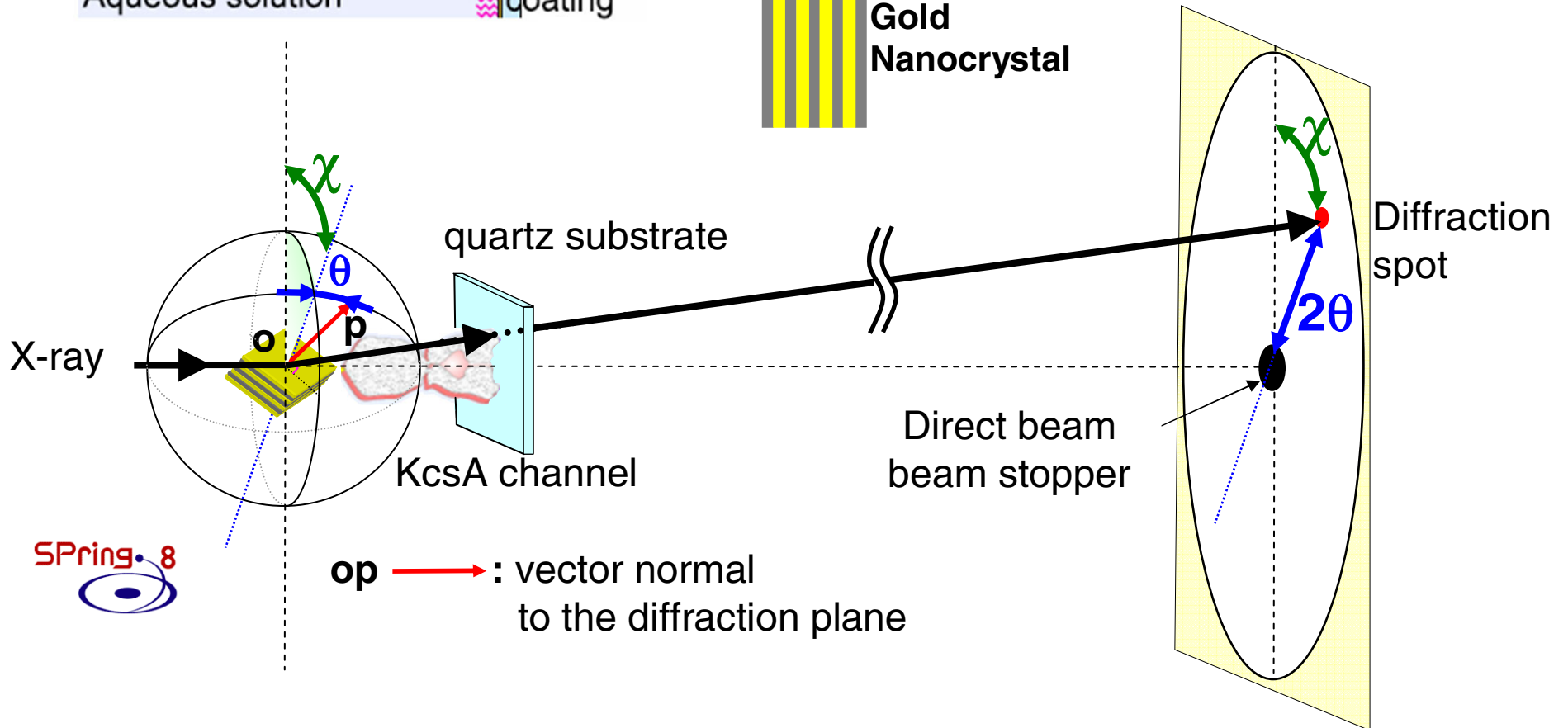
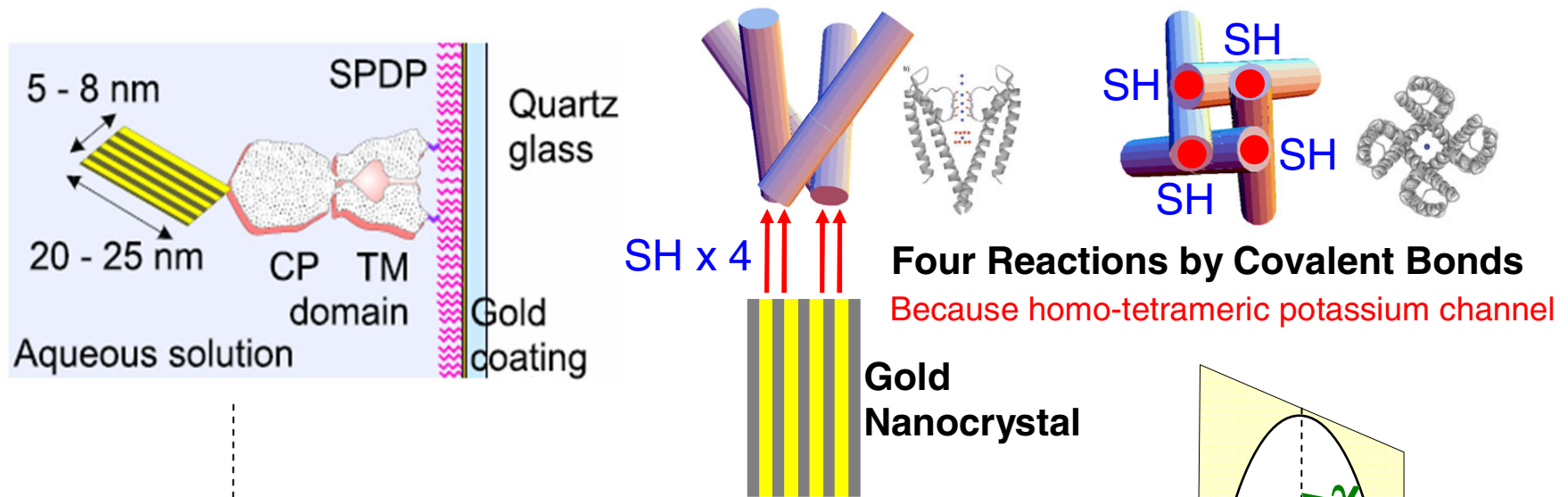


The patch clamp technique:

1991 Nobel Prize
(Prof. E. Neher and Prof. B. Sakmann)

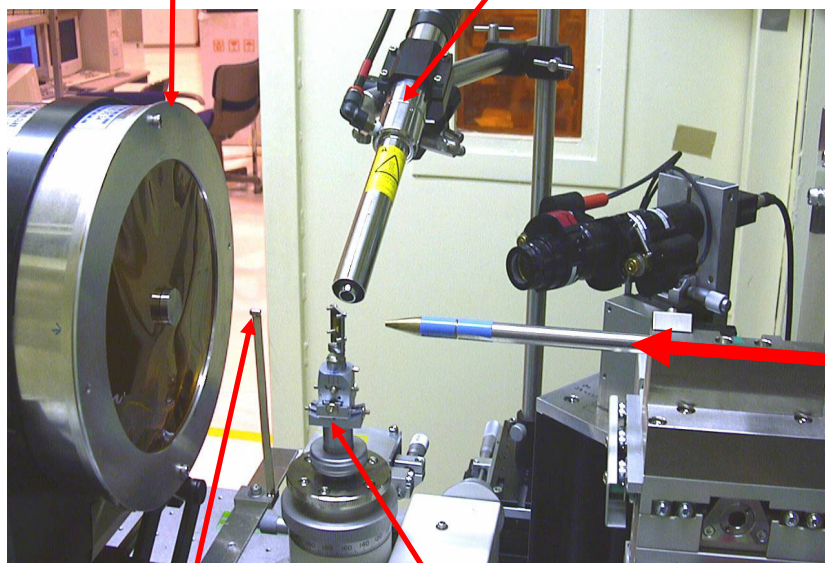


Arrangements of Adsorbed KcsA and DXT



Instrumentation:

Image Intensifier **Cooling System**



Direct Beam Stopper **Sample Holder**

BL44B2 (White X-ray Mode)

at SPring-8 (photon flux = 10^{15} photon/sec/mm²)

$\lambda = 1.7 \text{ \AA} (7 \text{ KeV}) - 0.4 \text{ \AA} (30 \text{ KeV})$

Detector: X-ray Image Intensifier (Hamamatsu, V5445P) with CCD

Specimen-Detector Distance: 10cm

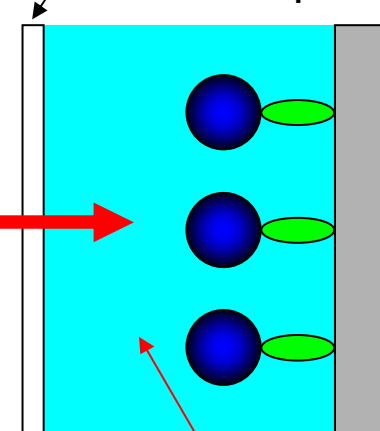
Exposure time: **within 1 second** (1-20 msec. x 33)

Cross-sectional view of the sample in DXT:

Polyimide film (7.5 μm)

quartz (70 μm)

X-ray (SR)
($\phi = 200 \mu\text{m}$)



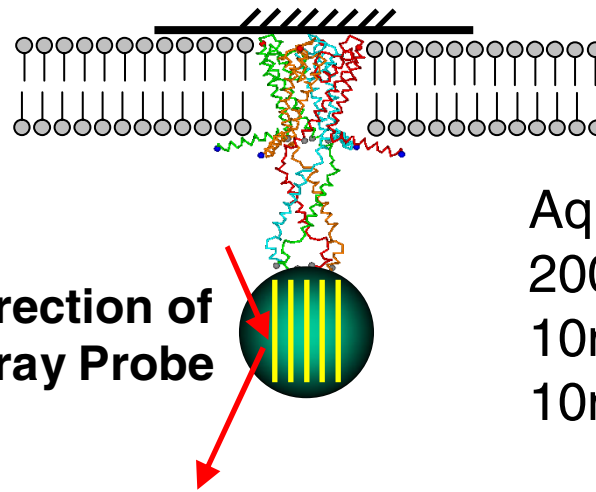
Aqueous Solution (7~10 μm)

 : gold nanocrystal

 : molecule

**The important thing:
The diffraction spots must not overlap in one Laue image to distinguish the observed diffraction spots**

DXT images:



Diameter of Nanocrystal:
20-30nm

during 1s, Image Speed = 1/5

Aqueous Solution:

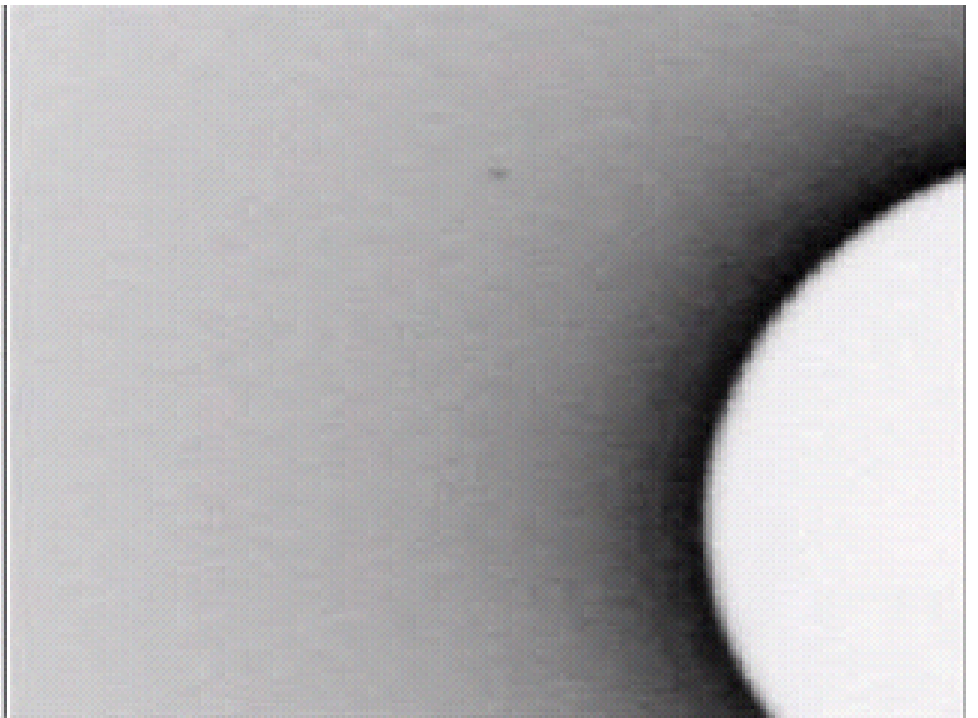
200mM KCl+

10mM HEPES for pH=7.5

10mM Succinic Acid for pH=4.0

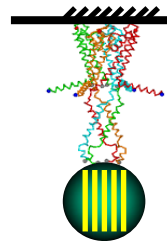
pH=7.5 (closed state)

pH=4.0 (gating state)

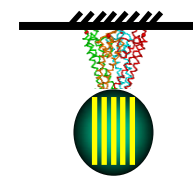


Histograms and MSD curves

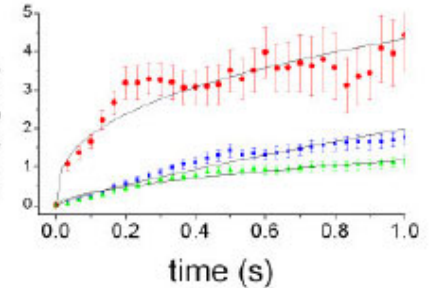
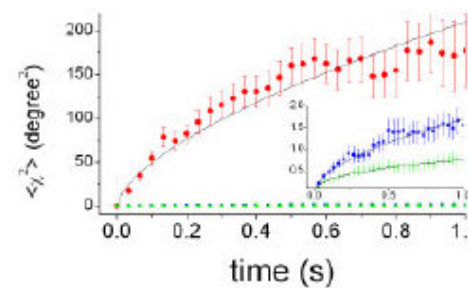
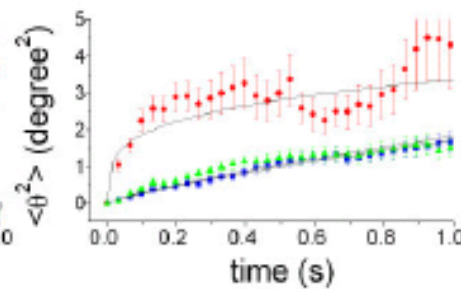
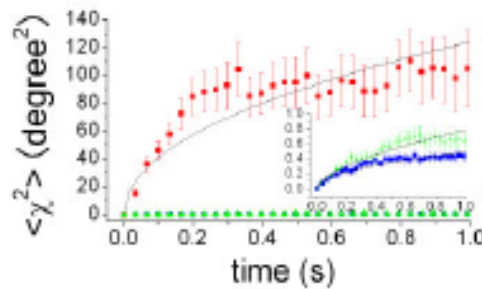
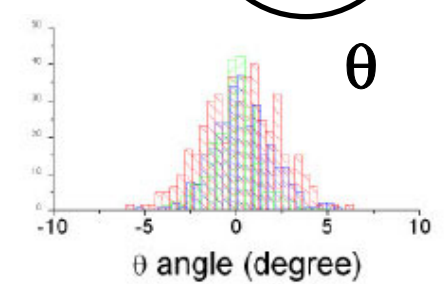
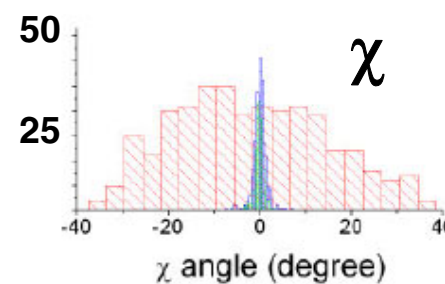
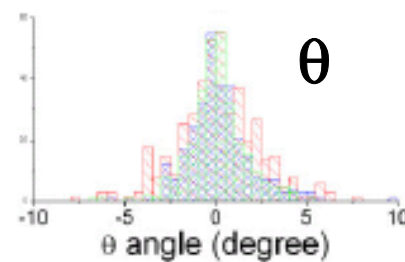
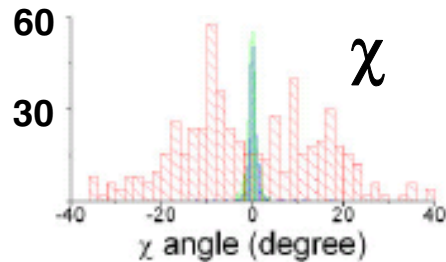
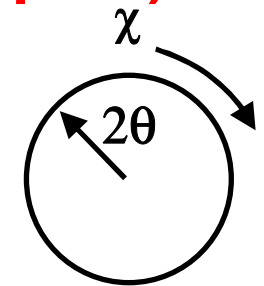
(from 400 spots)



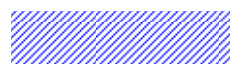
Full length KcsA



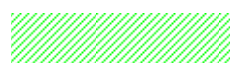
CP domain-deleted KcsA



: pH=4.0 (gating state)

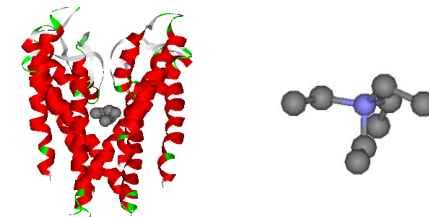


: pH=7.5 (closed state)



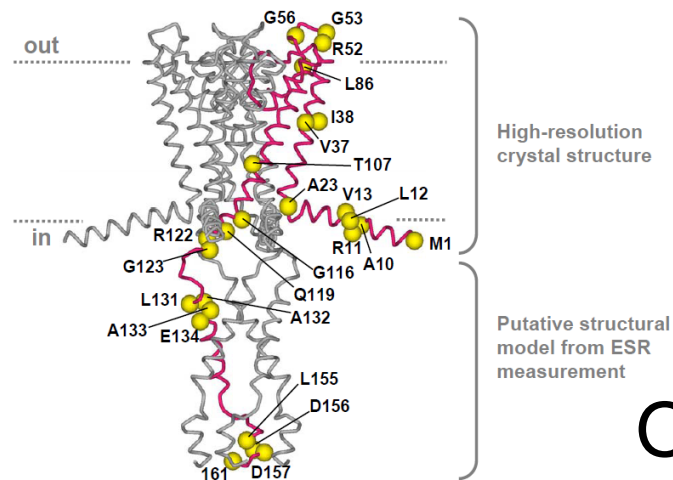
: pH=4.0 + TBA (open-locked state)

Position of Channel blocker



TBA (tetrabutylammonium)

Other data (KcsA)

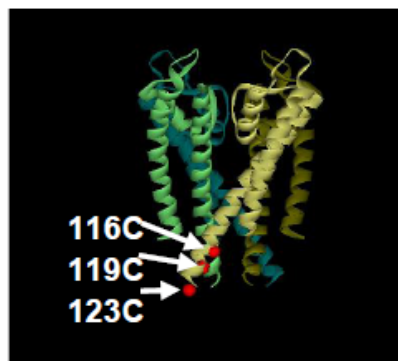


High-resolution crystal structure

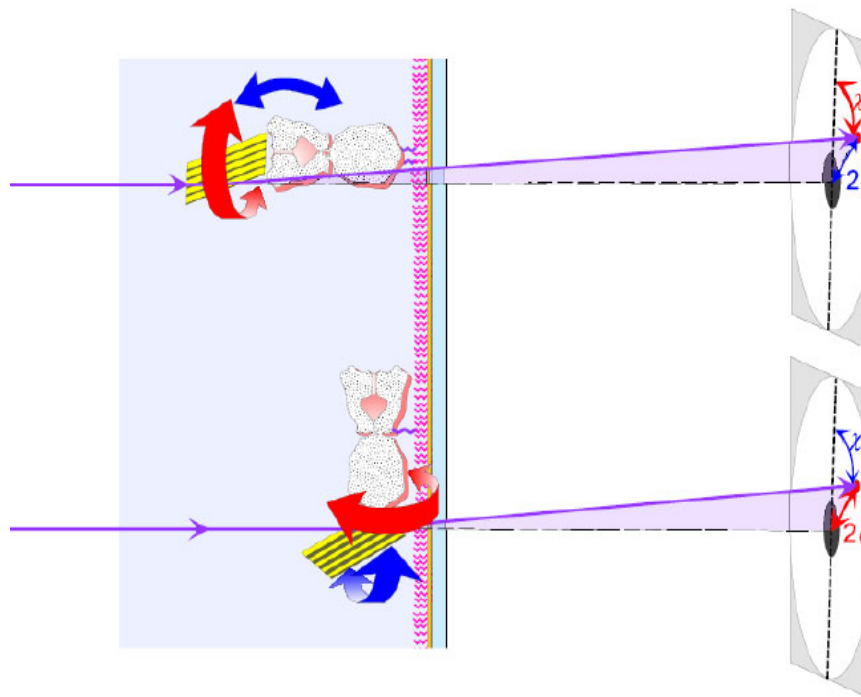
Putative structural model from ESR measurement

We utilized about twenty kinds of mutants of KcsA molecules in this experiment.

Other orientations of adsorbed KcsA



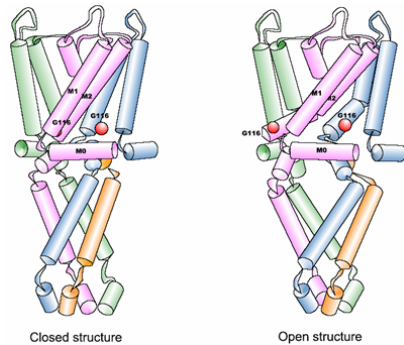
Other labeling sites



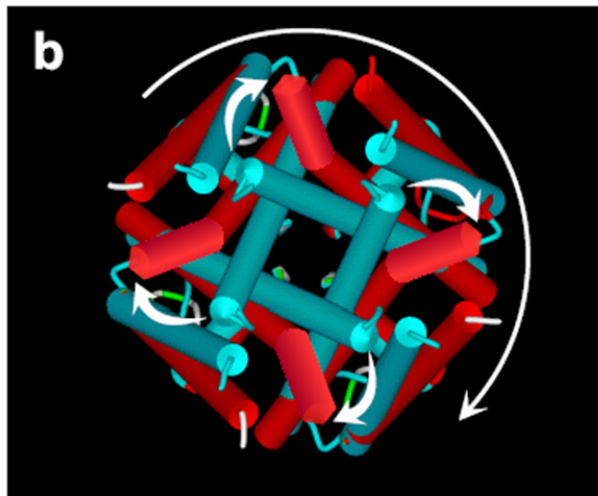
Twisting motions upon gating of KcsA

Transmembrane domain of KcsA

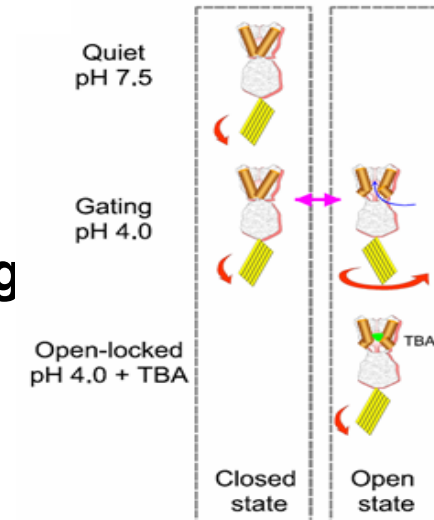
From structural data of KcsA crystals for opened and closed states, additionally, our dynamical observations, we can show the gating model.



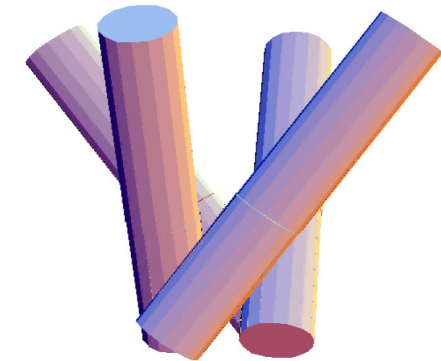
Open direction upon gating



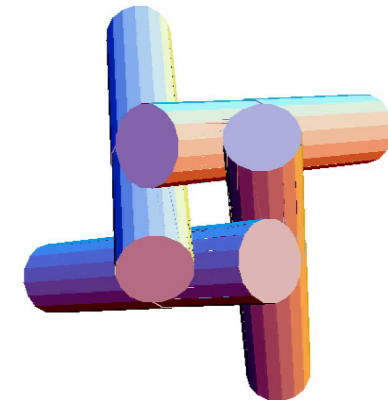
View from CP domain (intracellular side)



teal closed
red opened

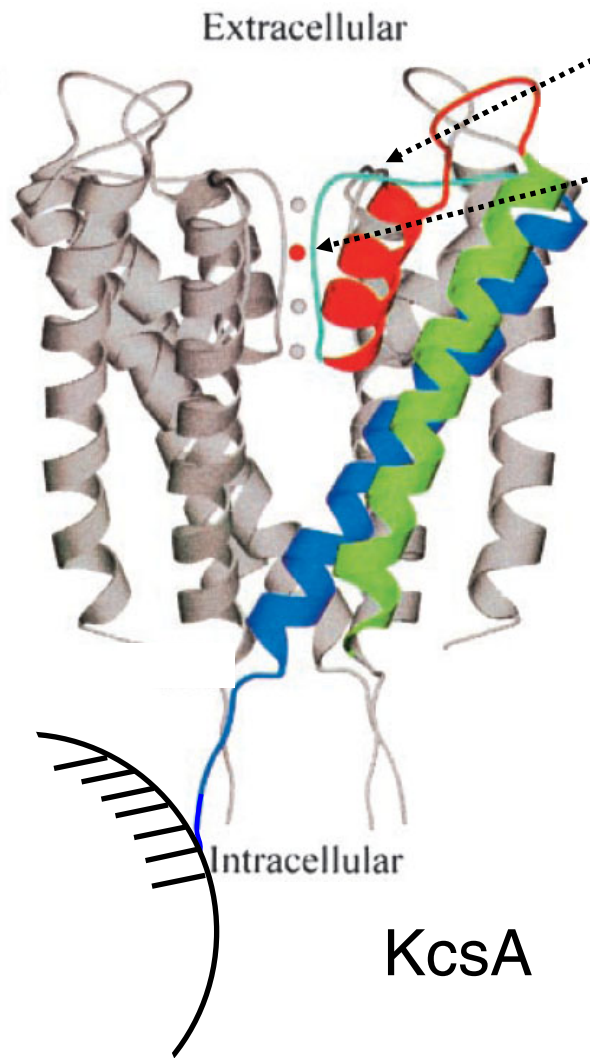


from side view



from CP domain side

Next observations (KcsA, KvAP, etc.,)



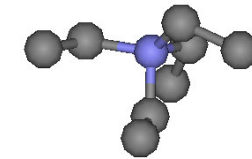
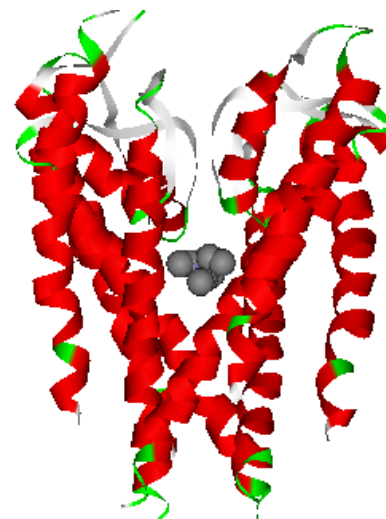
(a) Observations of extracellular domain.

(b) Fast observations of filter part.

(c) Simultaneous observations.

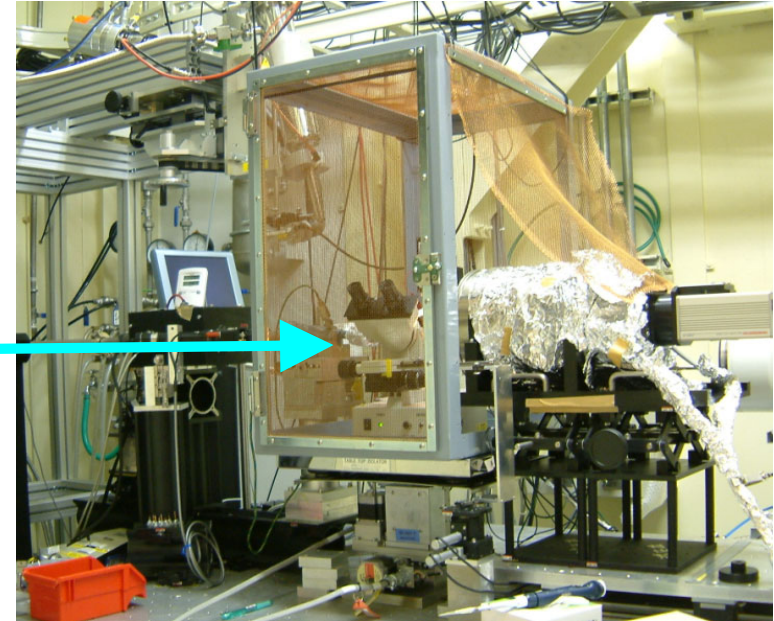
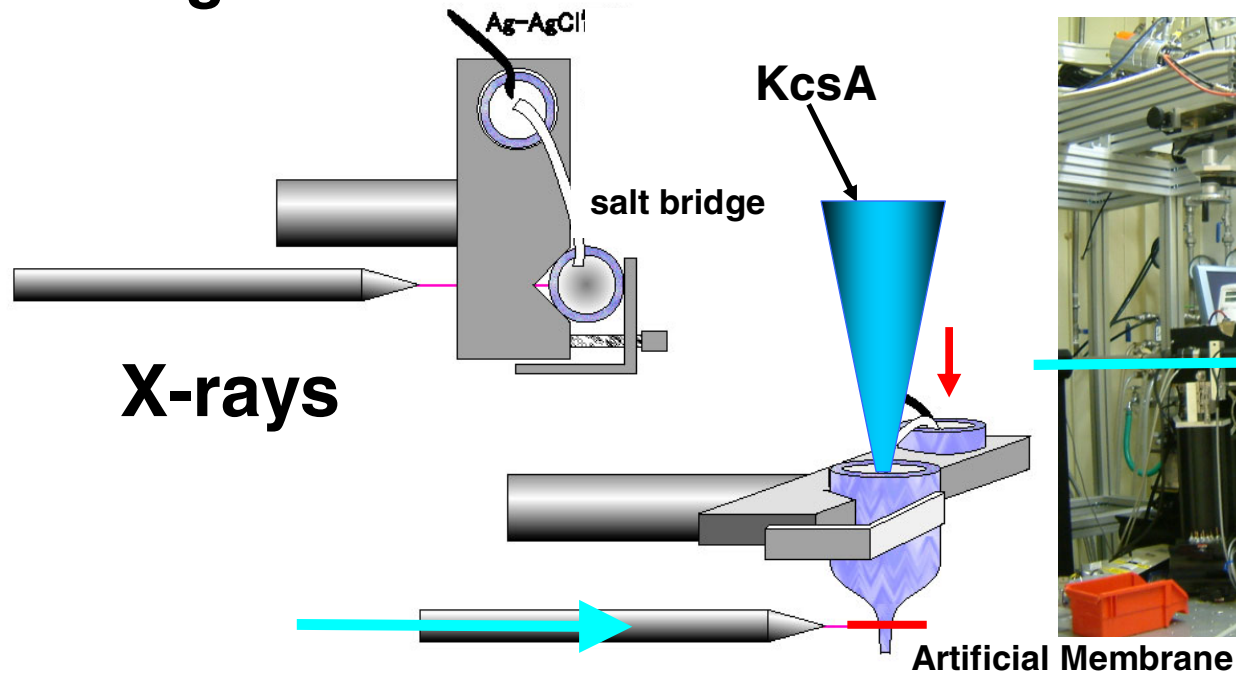
(d) *In-vivo* observations.

Position of Channel blocker

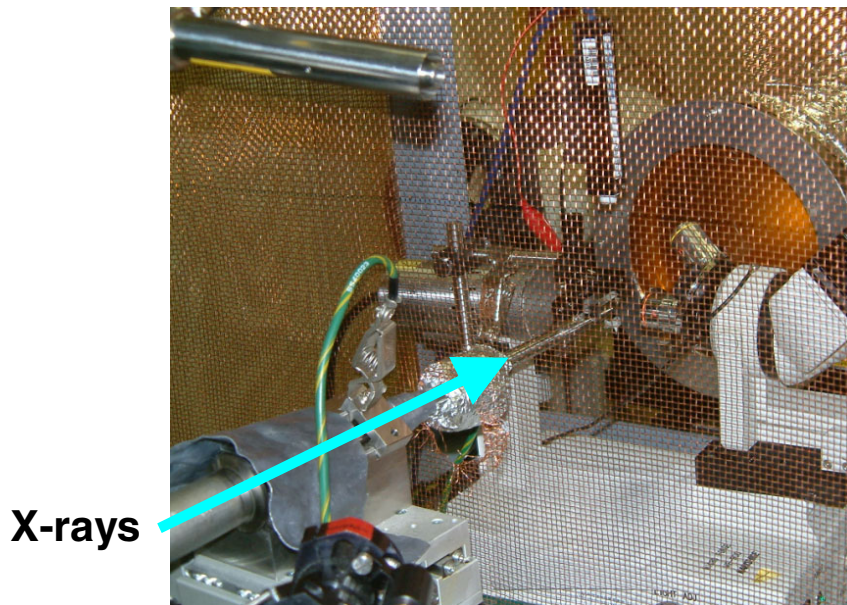


TBA (tetrabutylammonium)

Arrangement of Simultaneous Observations

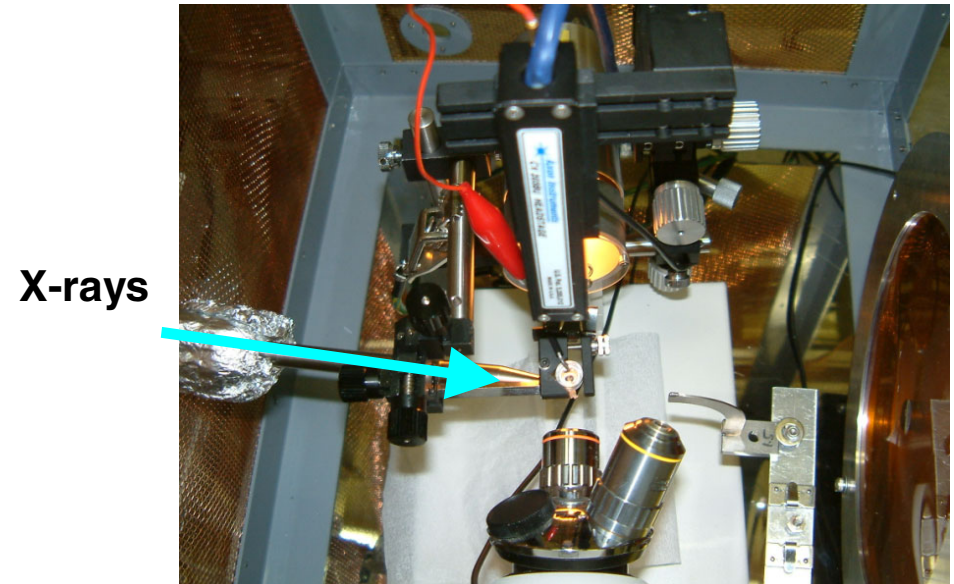


Artificial Membrane



X-rays

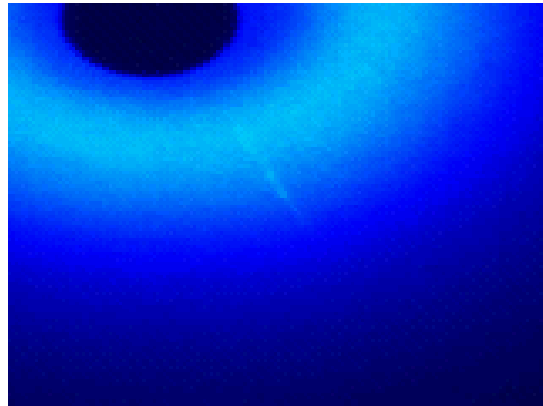
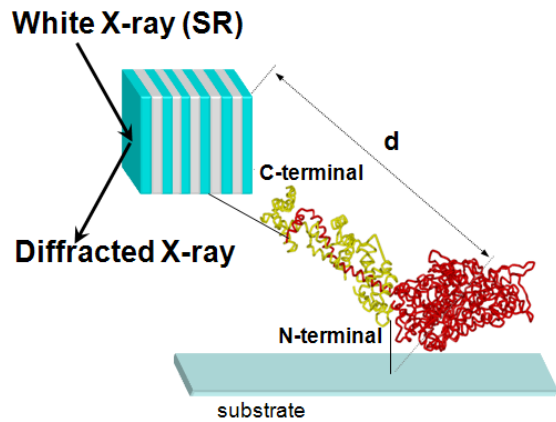
Front View



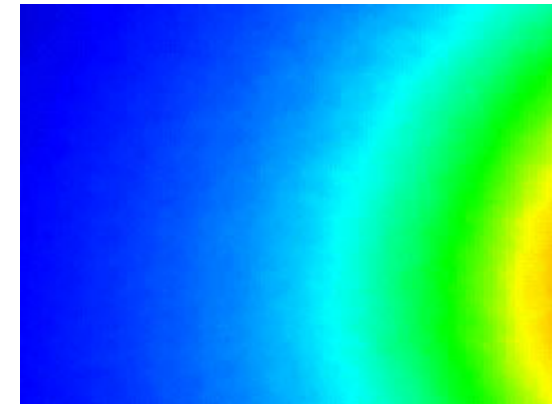
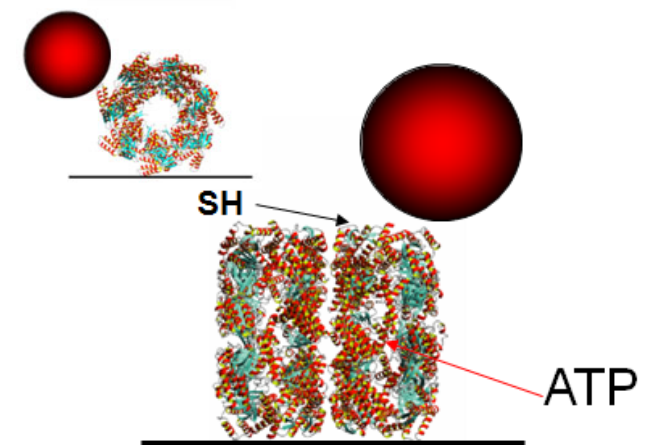
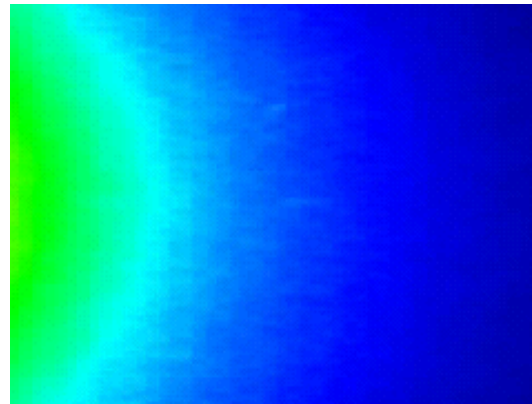
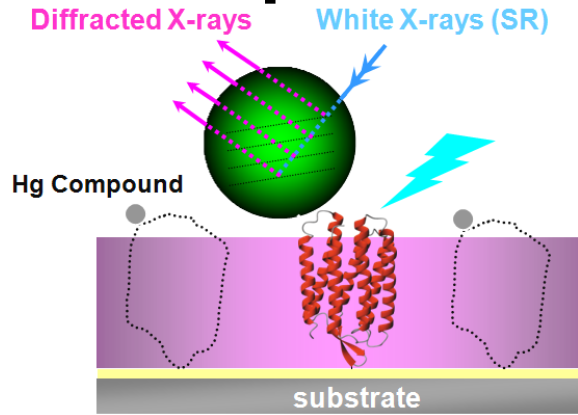
X-rays

Upper View

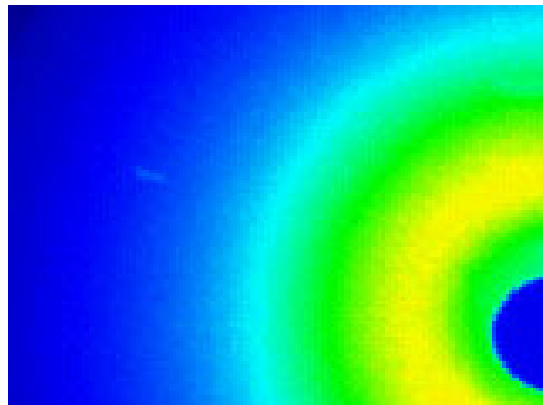
Other Experimental Samples:



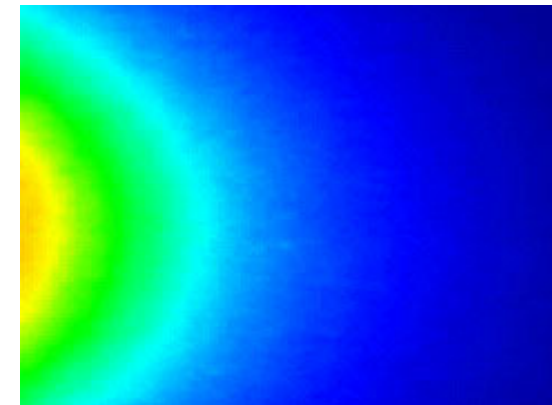
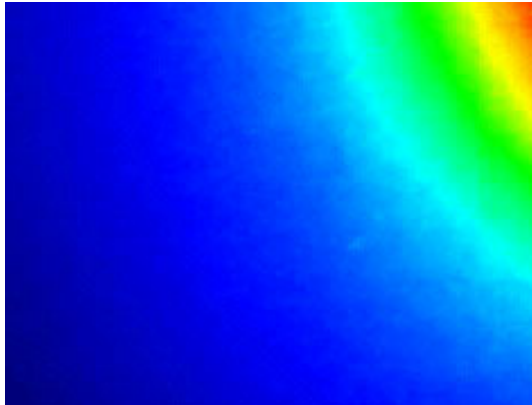
Mg-ATP(2mM)



Without ATP(2mM)



Ca-ATP(2mM)



With ATP(2mM)

Next targets for DXT:

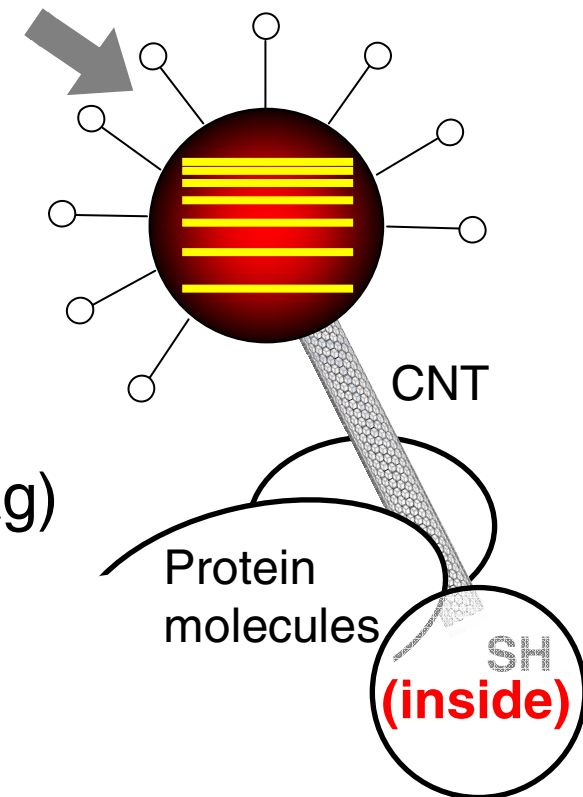
- (1) Simultaneous observations of single molecule (KcsA, KvAP, AChR).
- (2) *In vivo* observations.
- (3) High speed observations ($-\mu\text{s}$) using modified fast CCD.
- (4) DXT using monochromatic beam. We need **multi-d-space nanoparticles**.
- (5) Fabrications of magnetic nanocrystal and other nanocrystal.
(Fe_3O_4 , Co, CdSe, Ag)
- (6) Application of X-ray radiation pressure force to new microscope.
- (7) Application to Electron microscopy.
(EBSP or EBSD).

Still Photographs
(from crystal data)



**Real-time Movie
From Single Molecules**

NanoProbe



$$2d\sin\theta = n\lambda$$

Scanning Electron Microscope (JEOL JSM 7000F)



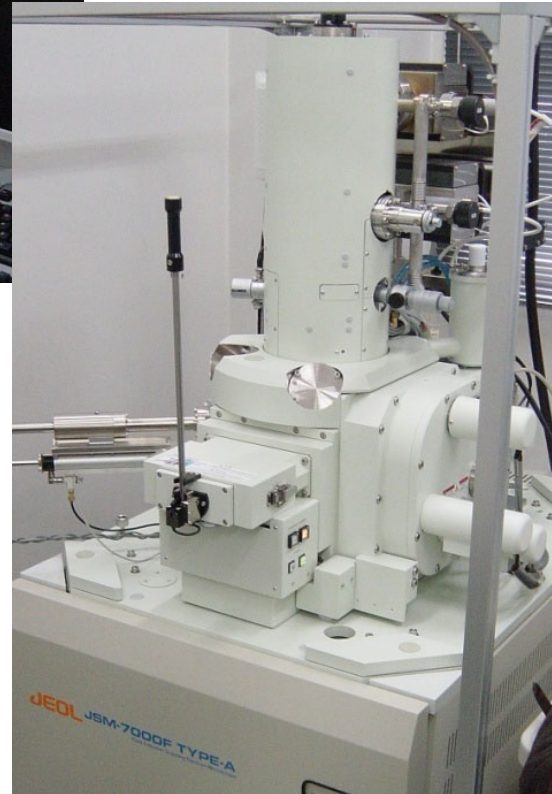
Laboratory Size

20cm

2km



Synchrotron Orbital Radiation Source (SPring-8, Japan)

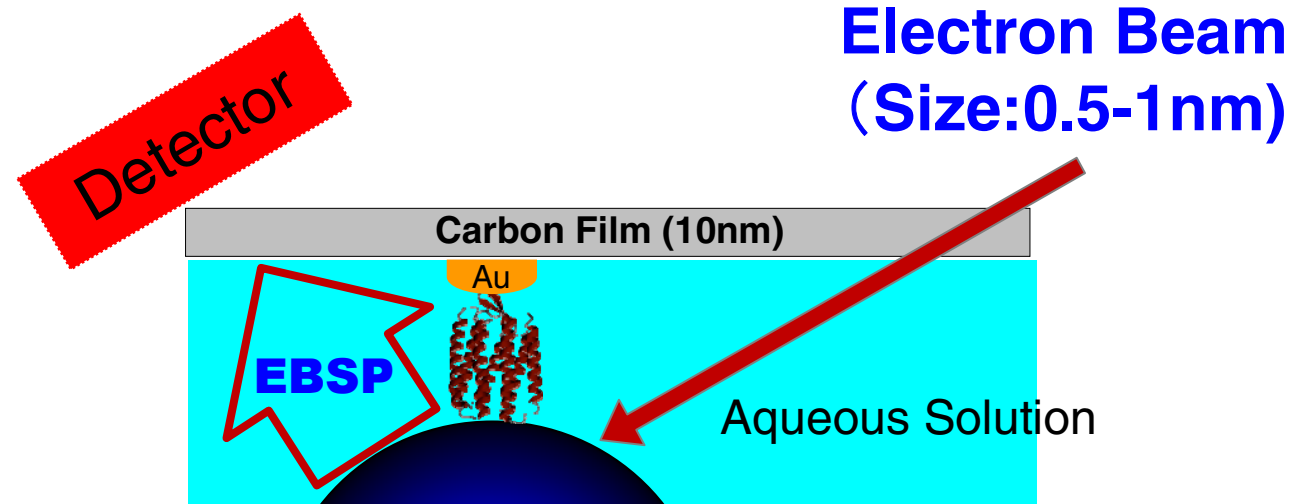


Comparison of X-ray and Electron Probe

	X-ray (DXT)	Electron (DET)
Accuracy	0.01 deg	0.1deg
Dimension	2D to 3D	3D
Label Size	20-60 nm	10-40 nm
Damage	Small	Very Large
Instrument	Large Institute	Lab. level
Machine Time	1 day/month	Everyday

Concept:

High speed detection before EBSP is attenuated



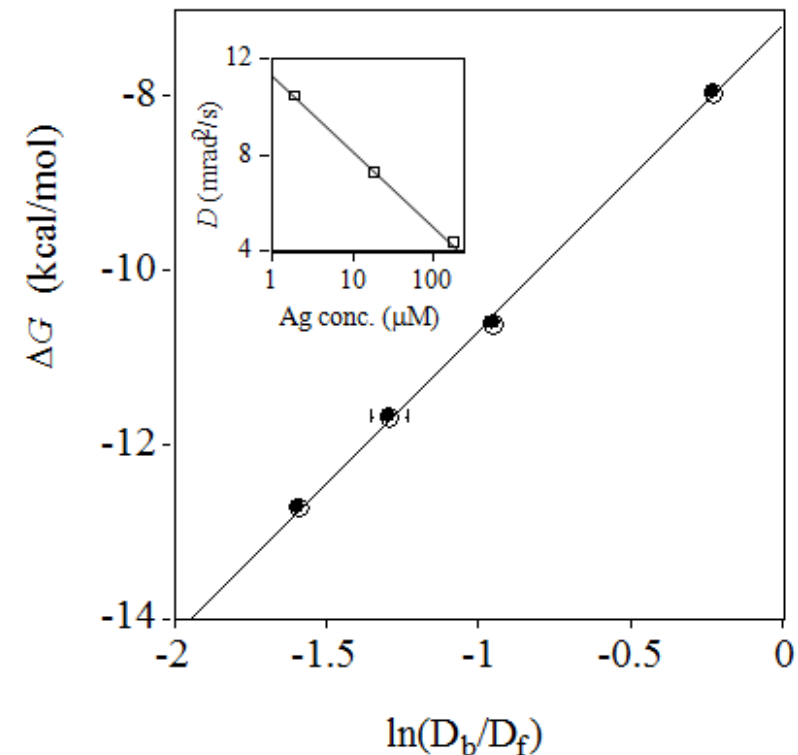
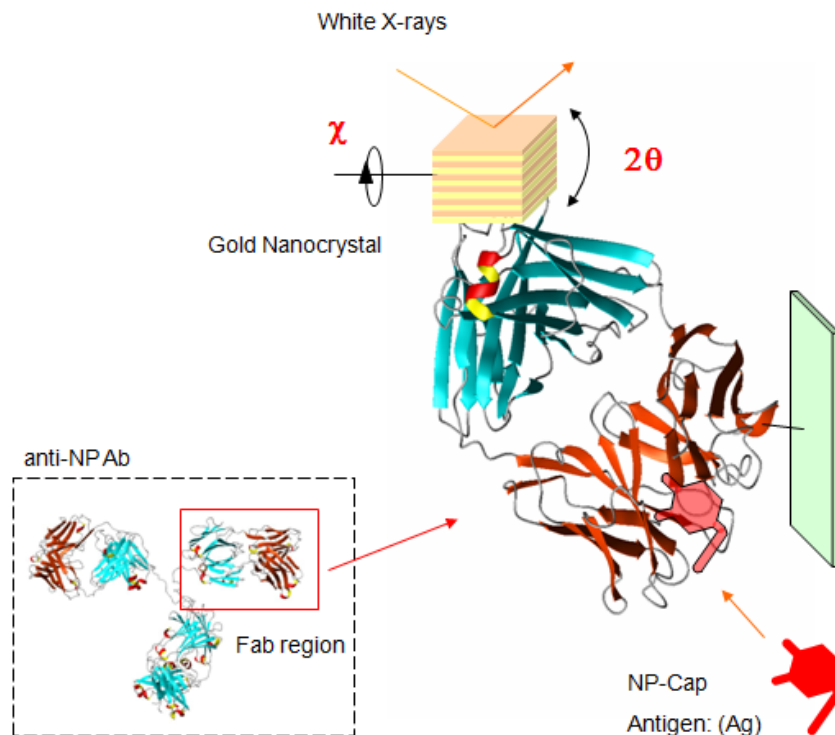
- (1) Carbon Thin Film(10nm) and Gold(10nm).
- (2) Pinpoint Electron Irradiation on Nanocrystal.

Mean free path (at 30KeV) :

Au ($\lambda=3\text{nm}$), C($\lambda=49\text{nm}$), H₂O($\lambda=84\text{nm}$)

Next Generation of Single Molecular World:

- (1) Structural Determination from Single Molecules
- (2) High Speed (ps-fs) and High Accuracy (pm)
- (3) Single Molecular Calorimeter
- (4) Control of Functional Dynamical Behaviors



A Special Thanks To:

SPring-8/JASRI Bio-soft Materials Group

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Teikyo University (Bio-engineering Center)

N. Ohishi

Nara Institute of Science and Technology (NAIST)

M. Kataoka

University College London (London Centre for Nanotechnology)

I. Robinson

Diamond Light Source

K. Inoue

CREST/JST

T. Oshima, T. Igarashi

The University of Tokyo

K. Ichiyanagi

KEK/Institute of Materials Structure Science

S. Adachi, S. Nozawa, T. Sato

